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Screening an Archetypal Collection of Microorganisms for the Presence of Unexplored Antimicrobial Compounds

Lucia Cimarelli[‡], Anna Maria Giuliodori[‡], Anna Brandi[‡], Karolina Adamkiewicz[§], Roberto Spurio[‡], Attilio Fabbretti[‡]

‡ University of Camerino, School of Biosciences and Veterinary Medicine, Camerino, Italy § Lodz University of Technology, Lodz, Poland

Corresponding author: Lucia Cimarelli (lucia.cimarelli@unicam.it)

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Abstract

Background

WHO estimates that more than 700.000 people die every year as a result of drug-resistant infections. To tackle this problem and change the current trend, it is necessary to design advanced strategies for drug discovery and to promote early-stage research activities finalized at the development of new drugs (World Health Organization 2015).

New information

In this study, we present the results of the preliminary screening of a library of microorganisms, collected from different environmental settings. Approximately 300 strains of the culture collection were tested on solid medium for inhibition of growth of three tester

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species, namely *Bacillus subtilis*, *Escherichia coli* and *Staphylococcus aureus*. The selection of the tester species was made according to the following criteria:

- Staphylococcus aureus and Escherichia coli were chosen as representative of Gram+ and Gram- bacteria, respectively. Furthermore, these organisms are relevant for the global public health because the number of antibiotic resistant strains responsible for invasive diseases is steadily increasing (Centers for Disease Control and Prevention 2014, Lowy 2003);
- *Bacillus subtilis*, instead, was selected as Gram+ tester microorganism commonly found in soil samples. Thus, this well characterized bacterium can be used to gain insight on the effect of metabolites produced by microorganisms of the culture collection described in this study.

One of the active strains, MES18, was classified as a *Bacillus* spp. by means of smallsubunit rRNA gene sequencing. To identify the compound(s) responsible for this inhibitory activity, MES18 cells were grown in liquid medium at 30°C and samples were taken at different time points over a period of 12 days. The supernatants obtained from the fermentation media were subjected to fractionation by chromatography on reversed-phase column and all the eluted compounds were assayed for their ability to repress the growth of tester strains. This approach allowed us to identify the fractions containing the bioactive compound(s) and to establish that the production of these secondary metabolites reached a maximum during the idiophase of the cell culture, when cell growth and replication decline. Further analyses to identify the physical-chemical features of the compound(s) produced by this strain using HPLC coupled to mass-spectrometry are currently ongoing.

Keywords

Drug-resistance, secondary metabolites, antibiotics.

Methods

In this study, more than 900 microorganisms were isolated from different environmental samples. They are maintained in an archetypal culture collection at the University of Camerino (Unicam). A Gram- (*Escherichia coli*) and two Gram+ microorganisms (*Bacillus subtilis* and *Staphylococcus aureus*) were used as tester species for the antibacterial activity investigation on approximately 300 strains of the culture collection. Each microorganism was streaked in a circle at the center of three plates of solid medium. Once the growth was markedly detectable, the three tester species were transferred on these plates by replica plating and the size of the inhibition zone was regularly monitored and measured. Subsequently, to identify the compound(s) responsible for the inhibitory activity, supernatants obtained from the fermentation media were taken at different time points over a period of about two weeks. These samples were subjected to fractionation by reverse-

phase chromatography and all the eluted compounds were evaluated by disk-diffusion assay.

Results and discussion

Using this straightforward approach, we identified several microorganisms able to produce secondary metabolites with antimicrobial activity. Here, we present the results of one of the active strains, MES18, isolated from a soil sample next to the Lake of Cingoli in Central Italy. Based on 16S rRNA sequence analysis, MES18 has been placed in the *Bacillus* spp. group. After five days of incubation, the inhibition displayed by MES18 reached 2mm in the case of *Escherichia coli*, 5mm in the case of *Staphylococcus aureus* and a notably reduced growth of *Bacillus subtilis*, which appears fuzzy over a zone of at least 25mm (Fig. 1). To investigate further this activity against *B. subtilis*, 20 ml of supernatant of MES18 cell culture, obtained after two days of incubation in LB medium at 30°C, were fractionated on reversed-phase column. This procedure allowed us to concentrate the metabolites, whose antibacterial activity displayed by fractions 3-5 is shown in Fig. 2. Further analysis aimed at identifying the physical-chemical properties of the compound(s) produced by MES18 (as well as of all the active microorganisms present in the culture collection) are currently ongoing. A combination of HPLC and mass-spectrometry techniques will be used to separate the bioactive compounds and to characterize their mass with high accuracy.

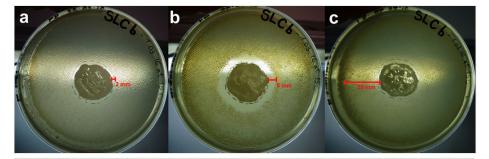


Figure 1.

Halo inhibition assay. LB agar plates with the active microorganism, MES18, tested on three tester species (**a** *E. coli*, **b** *S. aureus* and **c** *B. subtilis*). The inhibition zone is shown by a red line.

Conclusions

In this work, we have presented an investigation carried out on our collection of microorganisms (bacteria, fungi, algae) as part of an ongoing project on the identification of metabolites for bio-industrial use. The effect of metabolites produced by MES18 on three tester species is presented as a proof-of-concept study to highlight the potential repertoire of molecules synthesized by microorganisms. The isolation and characterization of these

active molecules is a key step to advance the research on new lead compounds that could be used as template for the development of new drugs. In light of the global challenge of the antibiotic-resistant infections, this research activity should be regarded as a 'categorical imperative' (in Kant's words).

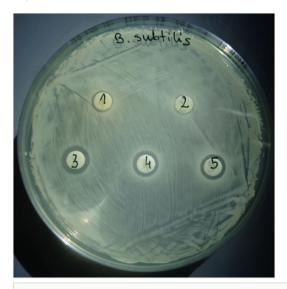


Figure 2.

Antimicrobial susceptibility test. Zones of inhibition of the tester strain *B. subtilis* were observed in three out of five disks containing fractions of reversed-phase column.

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Supplementary material

Suppl. material 1: Screening an archetypal collection of microorganisms for the presence of unexplored antimicrobial compounds

Authors: Lucia Cimarelli, Anna Maria Giuliodori, Anna Brandi, Karolina Adamkiewicz, Roberto Spurio, Attilio Fabbretti Data type: poster image Filename: Screening an Archetypal Collection of Microorganisms for the Presence of Unexplored Antimicrobial Compounds.pdf - <u>Download file</u> (395.82 kb)